

## Product datasheet

### AAV8 VP1, recombinant protein

#### Short overview

<b>Cat. No.</b>	640839
<b>Quantity</b>	10 µg
<b>Concentration</b>	100 µg/ml (1.19 µM)

#### Product description

<b>Formulation</b>	Liquid, 6 M urea in PBS
<b>Source</b>	Escherichia coli
<b>Molecular Weight</b>	83.9 kDa (calculated Mw from aa sequence)
<b>Purity</b>	> 95% (determined by SDS PAGE)
<b>Product description</b>	N-terminal His-tagged (MGSSHHHHHSSGLVPRGSH) recombinant AAV8 capsid protein VP1
<b>Purification</b>	Ni-NTA chromatography
<b>Storage</b>	-80°C
<b>Intended use</b>	Research use only
<b>Application</b>	Dot blot, SDS PAGE, WB

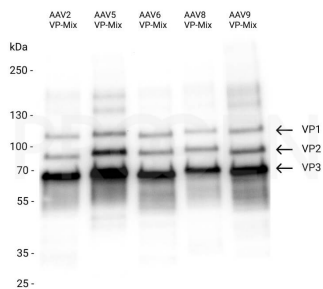
#### Applications

<b>Dot Blot</b>	100 ng, depending on primary antibody and detection method
<b>SDS PAGE</b>	1 µg
<b>Western Blot (WB)</b>	5-20 ng, depending on primary antibody and detection method

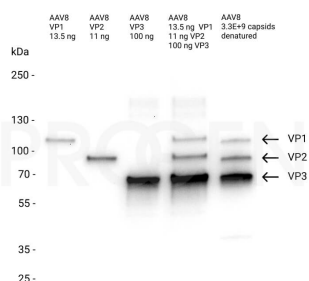
#### Background

The AAV capsid consists of three capsid proteins, i.e. VP1, VP2 and VP3, which differ in their N-terminus and encapsulate the genomic ssDNA. In native virus particles, the three proteins form subunits with a ratio of 1:1:10 (VP1:VP2:VP3), in a total number of 60 subunits per capsid. The recombinant AAV8 VP1 protein in combination with recombinant AAV8 VP2 (Cat. No. 640840) and recombinant AAV8 VP3 (Cat. No. 640841) can be used to create a mixture with the precise molar ratio of 1:1:10 to compare the protein composition of the viral capsid in your sample by protein detection methods, e.g. western blot. All three recombinant AAV8 capsid proteins are available as set (Cat. No. 72008) or as individual proteins (Cat. No. 640839, 640840, 640841). Note: please find an example how to prepare western blot samples in the pipetting scheme below. Aliquots of the remaining samples can be stored at -80°C for reuse.

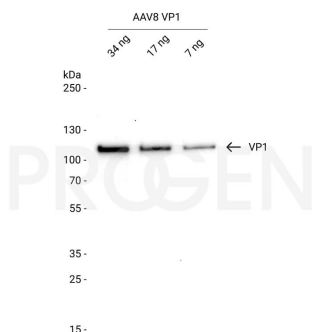
#### Product images



Western blot analysis of recombinant AAV2 VP proteins (Cat. No. 72001), recombinant AAV5 VP proteins (Cat. No. 72005), recombinant AAV6 VP proteins (Cat. No. 72006), recombinant AAV8 VP proteins (Cat. No. 72008) and recombinant AAV9 VP proteins (Cat. No. 72009) with B1 antibody (Cat. No. 690058). Western blot analysis was performed on the precise molar ratio of 1:1:10 (13.5 ng VP1:11 ng VP2:100 ng VP3) combined in one lane. The PVDF membrane was blocked with 5% milk in PBST for 1 h at RT. The primary antibody anti-AAV VP1/VP2/VP3, B1 (Cat. No. 690058) was diluted in blocking buffer (antibody concentration 500 ng/ml) and incubated for 1 h at RT. The secondary antibody anti-mouse IgG HRP was also diluted in blocking buffer (antibody concentration 200 ng/ml) and incubated for 1 h at RT. The bands were visualized by chemiluminescent detection using Pierce ECL Western Blotting Substrate.



Western blot analysis of recombinant AAV8 capsid proteins (Cat. No. 640839, 640840, 640841) and denatured AAV8 capsids with B1 antibody (Cat. No. 690058). Western blot analysis was performed on the precise molar ratio of 1:1:10 (VP1:VP2:VP3) either in separate lanes or combined in one lane and on 3.3E+09 denatured AAV8 capsids. The PVDF membrane was blocked with 5% milk in PBST for 1 h at RT. The primary antibody anti-AAV VP1/VP2/VP3, B1 (Cat. No. 690058) was diluted in blocking buffer (antibody concentration 500 ng/ml) and incubated for 1 h at RT. The secondary antibody anti-mouse IgG HRP was also diluted in blocking buffer (antibody concentration 200 ng/ml) and incubated for 1 h at RT. The bands were visualized by chemiluminescent detection using Pierce ECL Western Blotting Substrate.



Western blot analysis of recombinant AAV8 VP1 (Cat. No. 640839) with B1 antibody (Cat. No. 690058) and ECL detection. Western blot analysis was performed on different amounts of recombinant AAV-VP1 ranging from 7 ng to 34 ng. The PVDF membrane was blocked with 5% milk in PBST for 1 h at RT. The primary antibody anti-AAV VP1/VP2/VP3, B1 (Cat. No. 690058) was diluted in blocking buffer (antibody concentration 500 ng/ml) and incubated for 1 h at RT. The secondary antibody anti-mouse IgG HRP was also diluted in blocking buffer (antibody concentration 200 ng/ml) and incubated for 1 h at RT. The bands were visualized by chemiluminescent detection using Pierce ECL Western Blotting Substrate.