

Product datasheet

anti-AAV2, human chimeric, A20-h1

Short overview

Cat. No.	692379
Quantity	1 ml
Concentration	50 µg/ml

Product description

Host	Recombinant chimeric (human Fc region)
Antibody Type	Monoclonal
Isotype	IgG1
Clone	A20-h1
Immunogen	AAV2 capsids
Formulation	PBS, pH 7.4 with 0.09% sodium azide and 0.5% BSA
Binding affinity	KD value (AAV2) = 1.2E-10 M KD value (AAV3) = 9.3E-11 M
Synonym	Adeno-associated virus 2; AAV-2
Conjugate	Unconjugated
Purification	Affinity chromatography
Storage	2-8°C
Intended use	Research use only
Application	Dot blot, Neutralization assay, Serology ELISA
Reactivity	AAV2, AAV3
No reactivity	AAV1, AAV4, AAV5, AAV6, AAV8, AAV9, AAVDJ, AAVrh10, AAVrh74

Applications

Serology Assay	1-4 µg/ml (1:12.5-1:50; AAV2 ELISA)
Dot Blot	0.1 µg/ml (1:500; non-denaturing conditions)
Neutralization Assay	EC50 ~8 ng/ml (AAV2) and ~29 ng/ml (AAV3) - assay dependent

Background

Our human chimeric AAV antibodies are derived from our mouse monoclonal AAV antibodies and are a combination of the mouse antigen binding region and a human Fc region. Therefore, it provides the well-known characteristics of the corresponding mouse monoclonal antibody (anti-AAV2, A20, Cat. No. 61055) like cross-reactivity and neutralization activity combined with a human Fc region allowing the use in an anti-human secondary antibody detection system.

Many humans in the general population have developed antibodies against AAV as a result of naturally acquired infections, which might affect efficacy and safety of the gene transfer using AAV vectors. Therefore, testing for pre-existing AAV antibodies in patient sera is an indispensable step for the selection of patients for AAV gene therapy clinical trials.

Our human chimeric AAV antibodies are close to the human derived samples, making them the ideal positive control for reproducible and

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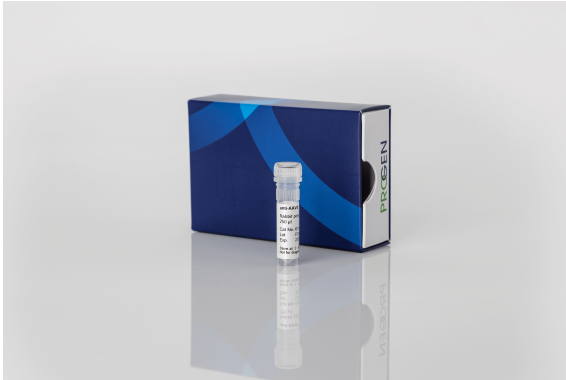
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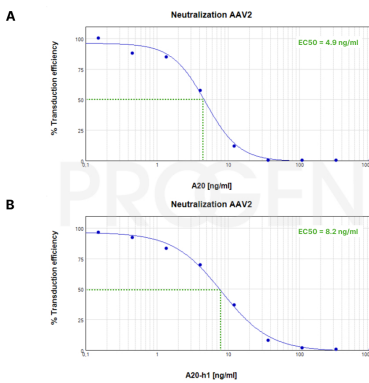
comparable serological assays.

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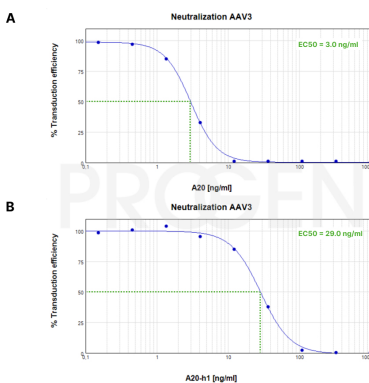
Product images



anti-AAV2, human chimeric, A20-h1



Neutralization of AAV2 with mouse monoclonal AAV2 antibody clone A20 (A) and human chimeric AAV2 antibody clone A20-h1 (B) by using AAV2-NanoLuc[®] viral particles from Promega. (A) anti-AAV2, mouse monoclonal, A20 or (B) anti-AAV2, human chimeric, A20-h1 were preincubated with AAV2-NanoLuc[®] viral particles for 30 min at RT at 300 rpm (antibody concentrations 0.2-3,000 ng/ml). HEK293 cells (100 μ l) were plated at 200,000 cells/ml in DMEM + 1% FCS. Virus-antibody-mix (20 μ l) was added to the cells and incubated for 16-24 h at 37°C. Extracellular NanoLuc Inhibitor and Nano-Glo[®] Live Cell Assay System (Promega) was added to the wells and incubated for 5 min at RT at 300 rpm. Luminescence was measured using an ID5-Reader and plotted with Softmax Pro 7.1 software to determine the EC50 values.



Neutralization of AAV3 with mouse monoclonal AAV2 antibody clone A20 (A) and human chimeric AAV2 antibody clone A20-h1 (B) by using AAV3-NanoLuc[®] viral particles from Promega. (A) anti-AAV2, mouse monoclonal, A20 or (B) anti-AAV2, human chimeric, A20-h1 were preincubated with AAV3-NanoLuc[®] viral particles for 30 min at RT at 300 rpm (antibody concentrations 0.2-3,000 ng/ml). HEK293 cells (100 μ l) were plated at 200,000 cells/ml in DMEM + 1% FCS. Virus-antibody-mix (20 μ l) was added to the cells and incubated for 16-24 h at 37°C. Extracellular NanoLuc Inhibitor and Nano-Glo[®] Live Cell Assay System (Promega) was added to the wells and incubated for 5 min at RT at 300 rpm. Luminescence was measured using an ID5-Reader and plotted with Softmax Pro 7.1 software to determine the EC50 values.